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## Genetic Diversity and Relationship of Selected Cultivars of Rice, *Oryza sativa* L. using Random Amplified Polymorphic DNA (RAPD) Markers

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### ABSTRACT

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The genetic diversity in 26 selected cultivars of rice, *Oryza sativa* L. using 20 decamer random primers. Out of 20, 15 RAPD primers revealed polymorphism while the remaining 5 primers showed no reaction. The primers produced a total of 71 bands of which 66 were polymorphic (95.2%). The number of polymorphic fragments for each primer varied from 1 to 3 with an average of 2.53 polymorphic fragments. The primer OPA-19 produced the maximum number of polymorphic bands. The RAPD data was analyzed to determine the genetic similarity coefficients which ranged from 0.46 to 0.81. Cluster analysis was performed using Ward's method) using the Jaccard's similarity coefficient. The Ward's method dendrogram resolved the selected rice cultivars into two to three major clusters.

### Introduction

The genus *Oryza* contains 25 recognized species, of which 23 are wild species and the remaining two are *O. sativa* and *O. glaberrima* which are cultivated species (Brar and Khush, 2003; Chang, 2003). *O. sativa* is the most widely grown worldwide including in Asian, North and South American, European Union, Middle Eastern and African countries. The world's rice production has doubled during last 25 years, largely due to the use of improved technology such as high yielding varieties and better crop management practices

(Byerlee, 1996). Demand for rice is growing every year and it is estimated that in 2025 AD the requirement would be 140 million tones. The land available for cultivation is decreasing due to continuous urbanization and inappropriate Rice (*Oryza sativa* L.) is one of the most important crops that provide food for more than half of the world population (Malik *et al.*, 2008). India has a long history of rice cultivation and stands first in rice area and second in rice production, after China. Approximately 90% of the world's rice is grown in the Asian

continent and constitutes a staple food for 2.7 billion people worldwide (Salim *et al.*, 2003; Paranthaman *et al.*, 2009).

Further scope of crop improvement depends on the conserved use of genetic variability and diversity in plant breeding programmes and use of new biotechnological tools. Molecular characterization can reveal the maximum genetic variation or genetic relatedness found in a population (Xu *et al.*, 2000). Chakravarthi and Naravaneni (2006) reported the usefulness of preservation and conservation of genetic resources since genetic diversity provides information to monitor germplasm and prediction of potential genetic gains. Information regarding genetic variability at molecular level could be used to help, identify and develop genetically unique germplasm that compliments existing cultivars (Ni *et al.*, 2002; Ravi *et al.*, 2003; Chakravarthi and Naravaneni, 2006). land use (Khush, 1997; Fischer *et al.*, 2000). To sustain present food self sufficiency and to meet future food requirements, India has to increase its rice productivity by 3 per cent per annum (Thiyagarajan and Selvaraju, 2001). DNA based molecular markers have proven to be powerful tools in the assessment of genetic variation and in the elucidation of genetic relationships within and among the species of rice (Ragunathanchari *et al.*, 1999, 2000; Shivapriya and Hittalmani, 2006). The present investigation was undertaken for the assessment of genetic diversity among the selected rice cultivars with the help of RAPD markers.

## **Materials and Methods**

Plant materials and genomic DNA isolation  
The plant materials selected for the present study were 26 different cultivars of rice. The seeds of 26 varieties such as Sakha 101, Sakha 102, Giza 172, Giza 178, Amber,

Furat, Yasmin, Mashkhab, Mashkhab, Branage-4, Al-abasia, Daawat, Aamchur, DPT-52, Gonor-1, Gundra, Guntur-2, Jeera Shankar, Komal, MTU, Patel-3, Shavagi, SHPP-20, Sonam, Soma, SS20, were collected from Rice Research Station, GKVK AND from farmer field of Karnataka, India. From traditional farmers of Karnataka, India. Healthy seeds of each variety were sowed in soil pots containing water under appropriate growth conditions for getting fresh leaves. DNA extraction was carried out from the fresh leaves collected from tillers following cetyl trimethyl ammonium bromide (CTAB) method (Doyle and Doyle, 1987) with some modifications. Freshly germinated 500 milligrams of young leaves were ground to a very fine powder in liquid nitrogen and dispersed in 3mL of pre-warmed (65°C) CTAB DNA extraction buffer (2% CTAB; 1.4 M NaCl; 100 mM Tris-HCl, pH 8.0; 20 mM EDTA, pH. 8.0; 0.2% - mercaptoethanol (added just before use). Oakridge tubes containing samples were incubated at 65°C for 30 min in a water bath. The samples were swirled every 10 min. After incubation the mixture was cooled down to room temperature and emulsified with an equal volume of chloroform: isoamyl alcohol (24:1) and centrifuged at 8000 rpm for 10 min. Following centrifugation, the aqueous phase was collected and nucleic acid was precipitated by mixing with equal volume of chilled isopropanol. The precipitated nucleic acid was centrifuged at 12000 rpm for 10 min and the pellet was washed with 70% ethanol. The DNA pellet obtained was dried and stored in 400 µL TE buffer.

## **Purification of DNA**

The RNA was removed by RNase treatment at 37°C for 30 min in a water bath. After incubation, DNA solution was extracted with an equal volume of chloroform:

isoamyl alcohol (24:1). The upper aqueous phase was collected after centrifugation at 8000 rpm for 10 min and mixed with 50  $\mu$ L of 3M sodium acetate. DNA was precipitated by adding two volumes of chilled absolute alcohol. The DNA pellet was air dried and dissolved in 100  $\mu$ L TE buffer. Two  $\mu$ L of genomic DNA isolated was subjected to electrophoresis on 0.8% agarose gel containing 1 mg/ mL ethidium bromide and the quantity of genomic DNA was assessed using undigested lambda DNA as control. For further use in PCR the DNA was diluted to a concentration of approximately 25 ng/ $\mu$ L.

### **RAPD analysis**

For the RAPD analysis of rice cultivars thirty deca-nucleotide primers of Operon Technology Inc. (Alameda, CA, USA) were used. The reaction was carried out in 25  $\mu$ L reaction volume containing 25 nanogram genomic DNA, 2.5  $\mu$ L 10X PCR buffer, 2  $\mu$ L 25 mM MgCl<sub>2</sub>, 2.5  $\mu$ L 2.5 mM dNTPs, 0.4  $\mu$ L Taq DNA polymerase and 2  $\mu$ L primer. All the reaction chemicals except primers were procured from M/s. Genei, Bangalore, India

### **RAPD amplification procedure**

Samples for amplification were carried out using the method stipulated by Williams *et al.* (1990) with some modifications of thermal cycles. Amplification was performed in a thermal cycler with an initial denaturation of 94o C for 5 min followed by 35 cycles which contains denaturation at 94o C for 1 min followed by annealing in which the annealing temperature was adjusted based on the T<sub>m</sub> value of each primers and finally extension at 72o C for 2 min. After 35 cycles, there was a final extension step at 72o C for 10 min. All the reactions were amplified in a Gradient Palmcycler (Corbett Research, CG-96, Australia). Each

amplification reaction for the screened primers was replicated two times individually with the same procedure in order to verify that the RAPD markers were reproducible and consistent.

### **Electrophoresis and visualization of RAPD products**

Amplified products were fractionated by 1.5% agarose gel in 1X TBE buffer (pH-8.0) at 100 V for 2 h and stained with ethidium bromide. 1kb DNA ladder was used as size marker. The gels were visualized under a UV transilluminator and documented using a digital camera. Total number of bands and number of polymorphic bands present in each cultivar was detected from the gels and scored manually. Each polymorphic band was considered as binary characters and was scored 1 (presence) or 0 (absence) for each sample. Only those fragments with medium and high intensity were taken into consideration.

### **Data analysis**

The gel images were scored using a binary scoring system that recorded the presence and absence of bands as “1” and “0” respectively. From the binary data, the similarity coefficient values between the cultivars were derived based on the probability that a particular character of one accession will also be present in another with the Jaccard’s correlation analysis using the statistical software. The statistical analysis is performed using Euclidean distance method statica 5.0 in order to cluster the accessions.

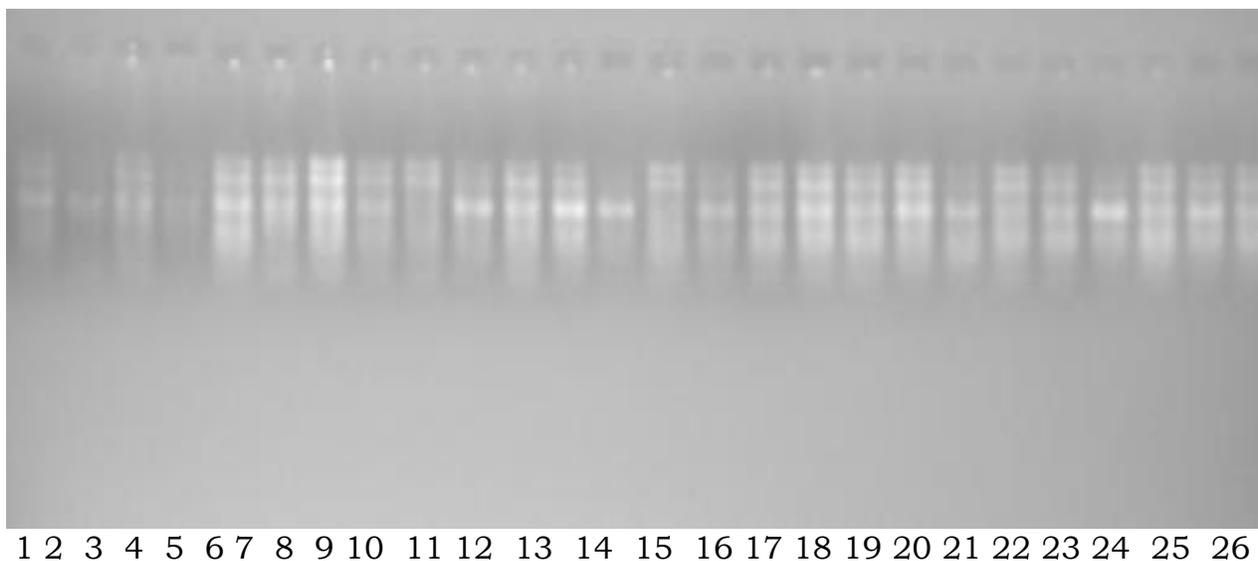
### **Results and Discussion**

The results of present study indicated a considerable level of genetic diversity among the cultivars selected. Among 20 primers used in this study, results of 15

primers were taken into consideration since they had given reproducible bands. Each polymorphic RAPD marker was considered as a locus so that every locus had two alleles, identified by the presence and absence of the band. A total of 428 DNA fragments were generated by 15 primers out of which 66 were polymorphic (95 % polymorphism) (Table 2). Out of 20 primers, only 15 primers exhibited more than 90% polymorphism. The number of polymorphic fragments for each primer varied from 65-71 with an average of 2.73 polymorphic fragments. The primer OPA-17 produced the maximum number of polymorphic bands. The percentage of polymorphism was calculated as 95%. The size of amplified fragments ranges between 300bp to 3000 bp (Figure 1). It was observed that the level of polymorphism with primers differed between the cultivars. Similarity between the cultivars was derived by Jaccard's

correlation coefficient (Jaccard, 1908). Correlation matrices obtained from all the primers used were consolidated in one single matrix and the mean values were presented (Table 3). Jaccard's pair-wise similarities computed between the cultivars showed that Sakha 101, Sakha 102, were the closest (0.91). The greatest distance was observed between the cultivars Daawat, and SS20, (0.46). RAPD data generated by twenty primers were subjected to Wards Euclidean distance cluster analysis and the dendrogram was constructed (Figure 2). Cluster analysis revealed the similarity between the rice cultivars and it ranged from 50% to 80%. The dendrogram classified the cultivars into two distinct clusters. The first cluster included three cultivars, Shavagi, SHPP-20, Sonam, . The second cluster included 21 cultivars collected from Farmers field of Tumkur .

**Fig.1** RAPD molecular markers in Rice varieties amplified by OPA-19



The present investigation revealed the effectiveness of RAPD in detecting polymorphism among different 26 cultivars of rice. The success of RAPD analysis in *O. sativa* accessions were also reported earlier (Muhammad *et al.*, 2005; Rahman *et al.*,

2007; Malik *et al.*, 2008). The percentage of polymorphism was found near to be 95 %. One of the reasons for this high level of polymorphism can be due to intraspecific variation among the cultivars. Information on intraspecific variation from the present

study might be useful in making decision for improvement of rice cultivars. Similarity level up to 80% in cluster analysis is indicative of plant derived from interspecific hybridization (Marsolais *et al.*, 1993). Six cultivars Aamchur, DPT-52, Gonor-1, Gundra, Guntur-2, Jeera Shankar, collected from Rice field Tumkur were grouped in a single cluster indicating more similarity among them and expressed more diversity from all the cultivars collected from traditional farmers of Tumkur . It is interesting to note that Giza 178, collected from farmers field of Tumkur was totally excluded from both the clusters. The findings of Reby Skaria *et al.* (2011) indicate that Gonor-1, Gundra, Guntur-2, are genetically distant.

The present findings confirm that genetic diversity of the plants is closely related to their geographic distribution. It has been reported that species with a wide geographic area generally have more genetic diversity (Wilkie *et al.*, 1993). The present investigation reveals that RAPD is a valuable tool for estimating the extent of genetic diversity as well as to ascertain the genetic relationship between different cultivars of *Oryza sativa*.

In conclusion, the present study revealed that the levels of genetic differentiation between cultivars of *O. sativa* increased with geographical distance. The polymorphism detected among 26 accessions will be helpful in selecting genetically diverse cultivars in future breeding programme. However, there were some precincts in the present study that only twenty six cultivars and twenty primers were used in RAPD analysis and hence reduce the chance to obtain a reliable knowledge precisely about the genetic structure of each cultivar of rice. Further studies involving large number of accessions and primers need to be conducted to get more precise

information.

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